

ENG

Instructions for Use:  
**HUMAN VEGF-C ELISA**

Catalogue number:  
**RAF138R**

**For research use only!**

 **BioVendor**  
**R&D**<sup>®</sup>



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## HISTORY OF CHANGES

Previous version	Current version
ENG.005.A	ENG.006.A
Product Data Sheet	Instructions for Use
The appearance of the document changed. (rebranding)	
History of changes added.	

### 1. INTENDED USE

The human VEGF-C ELISA is an enzyme-linked immunosorbent assay for the quantitative detection of human VEGF-C. The human VEGF-C ELISA is for research use only. Not for diagnostic or therapeutic procedures.

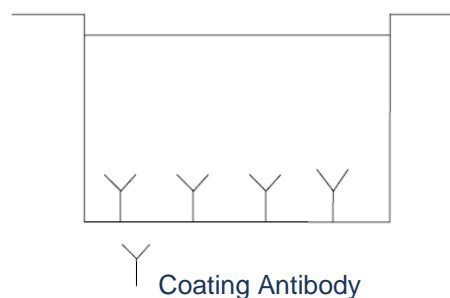
### 2. SUMMARY

Vascular endothelial growth factor-C (VEGF-C) is a member of the VEGF family of cytokines. It has been shown to exhibit angiogenic and lymphangiogenic actions. The VEGF family of growth factors and receptors is involved in the development and growth of the vascular endothelial system. Two of its family members, VEGF-C and VEGF-D, regulate the lymphatic endothelial cells via their receptor VEGFR3, thus acting as mitogens for these cells.

### 3. PRINCIPLES OF THE TEST

An anti-human VEGF-C coating antibody is adsorbed onto microwells.

*Figure 1 Coated Microwell*



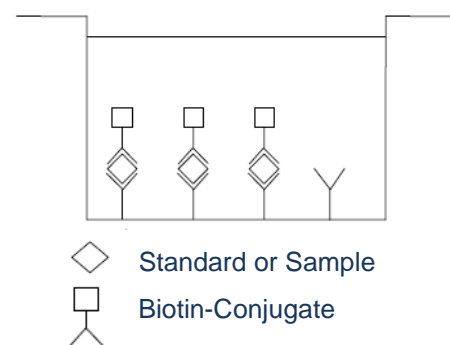
Human VEGF-C present in the sample or standard binds to antibodies adsorbed to the microwells.

*Figure 2 First Incubation*



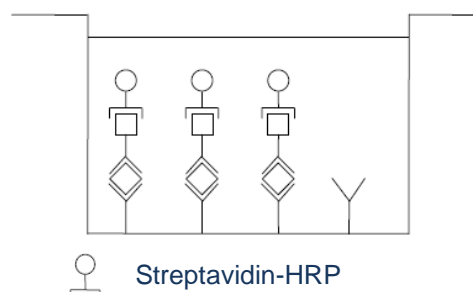
Following incubation unbound biological components are removed during a wash step and a biotin-conjugated anti-human VEGF-C antibody is added and binds to human VEGF-C captured by the first antibody.

*Figure 3 Second Incubation*



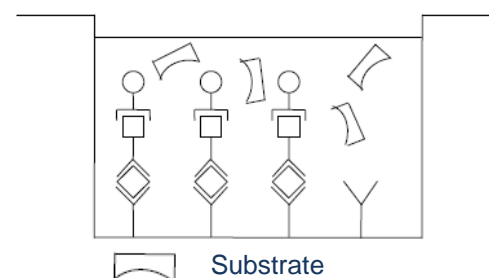
Following incubation unbound biotinconjugated anti-human VEGF-C antibody is removed during a wash step. StreptavidinHRP is added and binds to the biotinconjugated anti-human VEGF-C antibody.

*Figure 4 Third Incubation*



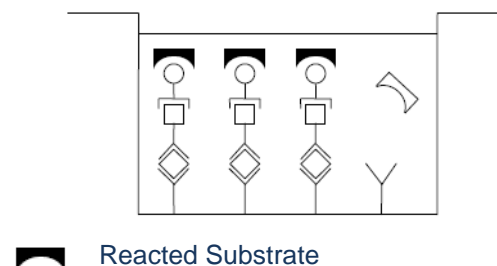
Following incubation unbound StreptavidinHRP is removed during a wash step, and substrate solution reactive with HRP is added to the wells.

*Figure 5 Fourth Incubation*



A coloured product is formed in proportion to the amount of human VEGF-C present in the sample or standard. The reaction is terminated by addition of acid and absorbance is measured at 450 nm. A standard curve is prepared from 7 human VEGF-C standard dilutions and human VEGF-C sample concentration determined.

Figure 6



#### 4. REAGENTS PROVIDED

- 1 aluminium pouch with a **Microwell Plate coated** with monoclonal antibody to human VEGF-C
- 1 vial (120 µl) **Biotin-Conjugate** anti-human VEGF-C monoclonal antibody
- 1 vial (120 µl) **Streptavidin-HRP**
- 2 vials human VEGF-C **Standard** lyophilized, 15 ng/ml upon reconstitution
- 1 vial (5 ml) **Sample Diluent**
- 1 vial (10 ml) **Assay Diluent**
- 1 vial (30 ml) **Conjugate Diluent**
- 1 bottle (50 ml) **Wash Buffer Concentrate** 20x (PBS with 1% Tween 20)
- 1 vial (15 ml) **Substrate Solution** (tetramethyl-benzidine)
- 1 vial (15 ml) **Stop Solution** (1M Phosphoric acid)
- 6 **Adhesive Films**

## 5. STORAGE INSTRUCTIONS – ELISA KIT

Store kit reagents between 2° and 8°C. Immediately after use remaining reagents should be returned to cold storage (2° to 8°C). Expiry of the kit and reagents is stated on labels. Expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, this reagent is not contaminated by the first handling.

## 6. SPECIMEN COLLECTION AND STORAGE INSTRUCTIONS

Cell culture supernatant, serum and plasma (citrate) were tested with this assay. Other biological samples might be suitable for use in the assay. Remove serum or plasma from the clot or cells as soon as possible after clotting and separation. Samples containing a visible precipitate must be clarified prior to use in the assay. Do not use grossly hemolyzed or lipemic specimens. Samples should be aliquoted and must be stored frozen at -20°C to avoid loss of bioactive human VEGF-C. Avoid repeated freeze–thaw cycles, preferentially use freshly collected samples. If samples are to be run within 24 hours, they may be stored at 2° to 8°C (for sample stability refer to 13.5). Prior to assay, the frozen sample should be brought to room temperature slowly and mixed gently. Do not thaw samples in a 37°C water bath. Do not vortex or sharply agitate samples.

## 7. MATERIALS REQUIRED BUT NOT PROVIDED

- 5 ml and 10 ml graduated pipettes
- 5 µl to 1000 µl adjustable single channel micropipettes with disposable tips
- 50 µl to 300 µl adjustable multichannel micropipette with disposable tips
- Multichannel micropipette reservoir
- Beakers, flasks, cylinders necessary for preparation of reagents
- Device for delivery of wash solution (multichannel wash bottle or automatic wash system)
- Microwell strip reader capable of reading at 450 nm (620 nm as optional reference wave length)
- Glass-distilled or deionized water
- Statistical calculator with program to perform regression analysis

## 8. PRECAUTIONS FOR USE

- All chemicals should be considered as potentially hazardous. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water. See material safety data sheet(s) and/or safety statement(s) for specific advice.
- Reagents are intended for research use only and are not for use in diagnostic or therapeutic procedures.
- Do not mix or substitute reagents with those from other lots or other sources.
- Do not use kit reagents beyond expiration date on label.
- Do not expose kit reagents to strong light during storage or incubation.
- Do not pipette by mouth.
- Do not eat or smoke in areas where kit reagents or samples are handled.
- Avoid contact of skin or mucous membranes with kit reagents or specimens.
- Rubber or disposable latex gloves should be worn while handling kit reagents or specimens.
- Avoid contact of substrate solution with oxidizing agents and metal.
- Avoid splashing or generation of aerosols.
- In order to avoid microbial contamination or cross-contamination of reagents or specimens which may invalidate the test use disposable pipette tips and/or pipettes.
- Use clean, dedicated reagent trays for dispensing the conjugate and substrate reagent.
- Exposure to acid inactivates the conjugate.
- Glass-distilled water or deionized water must be used for reagent preparation.
- Substrate solution must be at room temperature prior to use.
- Decontaminate and dispose specimens and all potentially contaminated materials as they could contain infectious agents. The preferred method of decontamination is autoclaving for a minimum of 1 hour at 121.5°C.
- Liquid wastes not containing acid and neutralized waste may be mixed with sodium hypochlorite in volumes such that the final mixture contains 1.0% sodium hypochlorite. Allow 30 minutes for effective decontamination. Liquid waste containing acid must be neutralized prior to the addition of sodium hypochlorite.

## 9. PREPARATION OF REAGENTS

**Buffer Concentrates** should be brought to room temperature and should be diluted before starting the test procedure. If crystals have formed in the **Buffer Concentrates**, warm them gently until they have completely dissolved.

### 9.1 Wash Buffer (1x)

Pour entire contents (50 ml) of the **Wash Buffer Concentrate** (20x) into a clean 1000 ml graduated cylinder. Bring to final volume of 1000 ml with glass-distilled or deionized water. Mix gently to avoid foaming. Transfer to a clean wash bottle and store at 2° to 25°C. Please note that Wash Buffer (1x) is stable for 30 days.

Wash Buffer (1x) may also be prepared as needed according to the following table:

Number of Strips	Assay Buffer Concentrate (20x) (ml)	Distilled Water (ml)
1 – 6	25	475
1 – 12	50	950

### 9.2 Biotin-Conjugate

Please note that the Biotin-Conjugate should be used within 30 minutes after dilution.

Make a 1:100 dilution of the concentrated Biotin-Conjugate solution with Conjugate Diluent in a clean plastic tube as needed according to the following table:

Number of Strips	Biotin-Conjugate (ml)	Conjugate Diluent (ml)
1 - 6	0.06	5.94
1 - 12	0.12	11.88

### 9.3 Streptavidin-HRP

Please note that the Streptavidin-HRP should be used within 30 minutes after dilution.

Make a 1:100 dilution of the concentrated Streptavidin-HRP solution with Conjugate Diluent in a clean plastic tube as needed according to the following table:

Number of Strips	Streptavidin-HRP (ml)	Conjugate Diluent (ml)
1 - 6	0.06	5.94
1 - 12	0.12	11.88



## 9.4 Human VEGF-C Standard

Reconstitute **human VEGF-C standard** by addition Sample Diluent (for subsequent measurement of **serum** or **plasma samples**) or Conjugate Diluent (for subsequent measurement of **cell culture supernatant samples**). Reconstitution volume is stated in the Quality Control Sheet. Swirl or mix gently to insure complete and homogeneous solubilization (concentration of reconstituted standard = 15 ng/ml).

Allow the standard to reconstitute for 10-30 minutes. Mix well prior to making dilutions. After usage remaining standard cannot be stored and has to be discarded.

### 9.4.1 Standard Dilution

Label 6 tubes, one for each standard point.

S2, S3, S4, S5, S6, S7

Then prepare 1:2 serial dilutions for the standard curve as follows:

Pipette 100  $\mu$ l of Sample Diluent (for subsequent measurement of serum and plasma samples) or Conjugate Diluent (for subsequent measurement of cell culture supernatant samples) into tubes S2 – S7.

Pipette 100  $\mu$ l of reconstituted standard (serves as highest standard S1 = 15 ng/ml) into the first tube, labelled S2, and mix (concentration of standard 2 = 7.5 ng/ml).

Pipette 100  $\mu$ l of this dilution into the second tube, labelled S3, and mix thoroughly before the next transfer. Repeat serial dilutions 4 more times thus creating the points of the standard curve (see Figure 7).

Sample Diluent (serum or plasma samples) or Conjugate Diluent (cell culture supernatant samples) serve as blank.

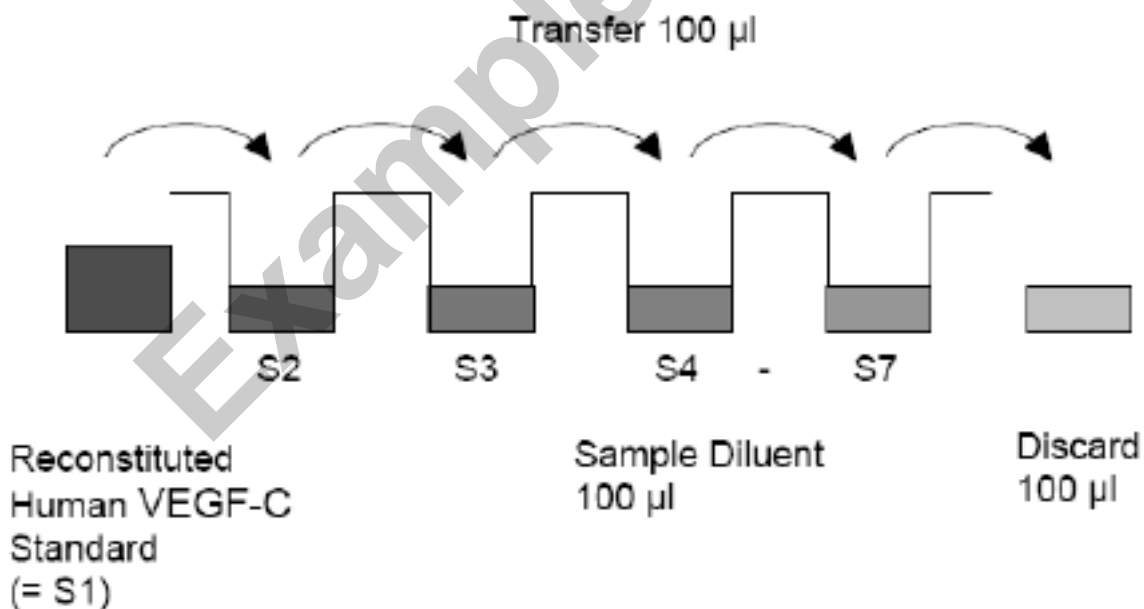


Figure 7

## 10. TEST PROTOCOL

- a) Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running blanks and standards. Each sample, standard, blank and optional control sample should be assayed in duplicate. Remove extra microwell strips from holder and store in foil bag with the desiccant provided at 2°-8°C sealed tightly.
- b) Wash the microwell strips twice with approximately 400 µl **Wash Buffer** per well with thorough aspiration of microwell contents between washes. Allow the Wash Buffer to sit in the wells for about **10 – 15 seconds** before aspiration. Take care not to scratch the surface of the microwells. After the last wash step, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess Wash Buffer. Use the microwell strips immediately after washing. Alternatively microwell strips can be placed upside down on a wet absorbent paper for not longer than 15 minutes. **Do not allow wells to dry.**
- c) Add 80 µl of **Assay Diluent** in duplicate to **all wells**.
- d) Add 20 µl of each prepared **Standard dilution** in duplicate to the corresponding **standard well** (see Table 1).
- e) Add 20 µl of each **sample** in duplicate to the **sample wells** (see Table 1).
- f) Add 20 µl of **Sample Diluent** (for serum or plasma samples) or **Conjugate Diluent** (for cell culture supernatant samples) in duplicate to the **blank wells**.

Table depicting an example of the arrangement of blanks, standards and samples in the microwell strips:

	1	2	3	4
A	Standard 1 (15.00 ng/ml)	Standard 1 (15.00 ng/ml)	Sample 1	Sample 1
B	Standard 2 (7.50 ng/ml)	Standard 1 (7.50 ng/ml)	Sample 2	Sample 2
C	Standard 3 (3.75 ng/ml)	Standard 1 (3.75 ng/ml)	Sample 3	Sample 3
D	Standard 4 (1.88 ng/ml)	Standard 1 (1.88 ng/ml)	Sample 4	Sample 4
E	Standard 5 (0.94 ng/ml)	Standard 1 (0.94 ng/ml)	Sample 5	Sample 5
F	Standard 6 (0.47 ng/ml)	Standard 1 (0.47 ng/ml)	Sample 6	Sample 6
G	Standard 7 (0.23 ng/ml)	Standard 1 (0.23 ng/ml)	Sample 7	Sample 7
H	Blank	Blank	Sample 8	Sample 8

Table 1

- g) Cover with an adhesive film and incubate at room temperature (18 to 25°C) for 2 hours, if available on a microplate shaker.
- h) Prepare **Biotin-Conjugate** (see Preparation of Biotin-Conjugate 9.2).
- i) Remove adhesive film and empty wells. **Wash** microwell strips 6 times according to point b. of the test protocol. Proceed immediately to the next step.
- j) Add 100 µl of Biotin-Conjugate to all wells.
- k) Cover with an adhesive film and incubate at room temperature (18 to 25°C) for 1 hour, if available on a microplate shaker.
- l) Prepare Streptavidin-HRP (refer to Preparation of Streptavidin-HRP 9.3).
- m) Remove adhesive film and empty wells. Wash microwell strips 6 times according to point b. of the test protocol. Proceed immediately to the next step.
- n) Add 100 µl of diluted Streptavidin-HRP to all wells, including the blank wells.
- o) Cover with an adhesive film and incubate at room temperature (18° to 25°C) for 1 hour, if available on a microplate shaker.
- p) Remove adhesive film and empty wells. Wash microwell strips 6 times according to point b. of the test protocol. Proceed immediately to the next step.
- q) Pipette 100 µl of TMB Substrate Solution to all wells.
- r) Incubate the microwell strips at room temperature (18° to 25°C) for about 30 min. Avoid direct exposure to intense light. The colour development on the plate should be monitored and the substrate reaction stopped (see next point of this protocol) before positive wells are no longer properly recordable. Determination of the ideal time period for colour development has to be done individually for each assay. Standard has developed a dark blue colour. Alternatively the colour development can be monitored by the ELISA reader at 620 nm. The substrate reaction should be stopped as soon as Standard 1 has reached an OD of 0.9 – 0.95.
- s) Stop the enzyme reaction by quickly pipetting 100 µl of Stop Solution into each well. It is important that the Stop Solution is spread quickly and uniformly throughout the microwells to completely inactivate the enzyme. Results must be read immediately after the Stop Solution is added or within one hour if the microwell strips are stored at 2 - 8°C in the dark.
- t) Read absorbance of each microwell on a spectro-photometer usány 450 nm as the primary wave length (optionally 620 nm as the reference wave length; 610 nm to 650 nm is acceptable). Blank the plate reader according to the manufacturer's instructions by using the blank wells. Determine the absorbance of both the samples and the standards.

**Note:** In case of incubation without shaking the obtained O.D. values may be lower than indicated below. Nevertheless the results are still valid.

## 11. CALCULATION OF RESULTS

- Calculate the average absorbance values for each set of duplicate standards and samples. Duplicates should be within 20 per cent of the mean value.
- Create a standard curve by plotting the mean absorbance for each standard concentration on the ordinate against the human VEGF-C concentration on the abscissa. Draw a best fit curve through the points of the graph (a 5-parameter curve fit is recommended).
- To determine the concentration of circulating human VEGF-C for each sample, first find the mean absorbance value on the ordinate and extend a horizontal line to the standard curve. At the point of intersection, extend a vertical line to the abscissa and read the corresponding human VEGF-C concentration.
- **Calculation of samples with a concentration exceeding standard 1 may result in incorrect, low human VEGF-C levels. Such samples require further external predilution according to expected human VEGF-C values with Sample Diluent in order to precisely quantitate the actual human VEGF-C level.**
- It is suggested that each testing facility establishes a control Hamper of known human VEGF-C concentration and runs this additional control with each assay. If the values obtained are not within the expected range of the control, the assay results may be invalid.
- A representative standard curve is shown in Figure 8. This curve cannot be used to derive test results. Each laboratory must prepare a standard curve for each group of microwell strips assayed.

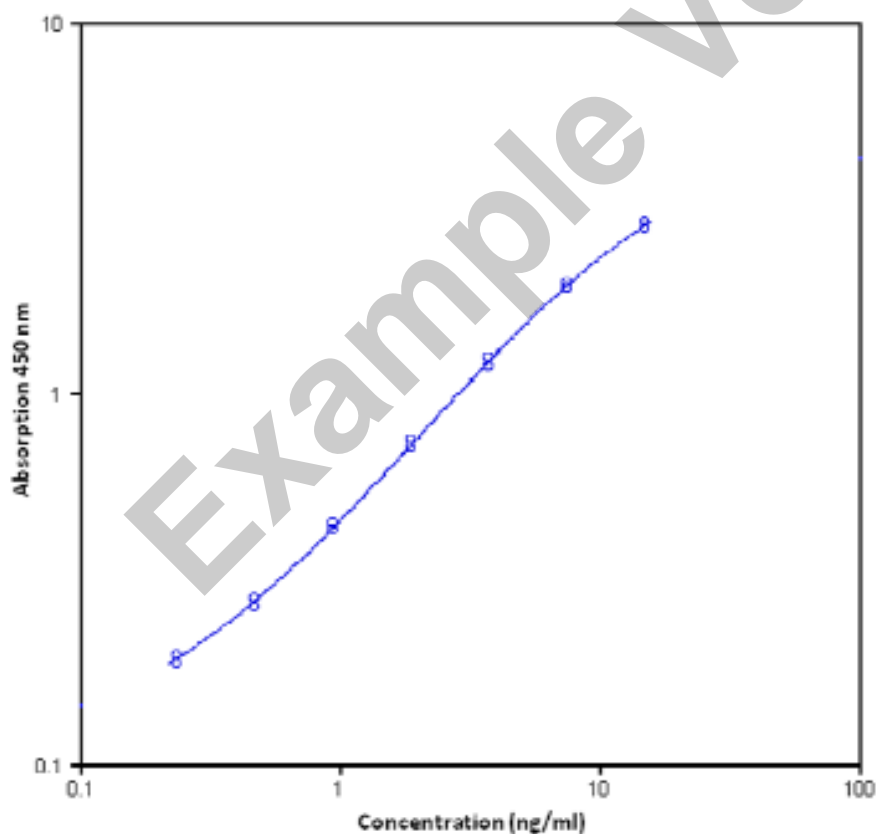


Figure 8

*Representative standard curve for human VEGF-C ELISA. Human VEGF-C was diluted in serial 2-fold steps in Sample Diluent. Do not use this standard curve to derive test results. A standard curve must be run for each group of microwell strips assayed.*

Typical data using the human VEGF-C ELISA  
 Measuring wavelength: 450 nm  
 Reference wavelength: 620 nm

Standard	Human VEGF-C Concentration (ng/ml)	O.D. at 450 nm	Mean O.D. at 450 nm	C.V. (%)
1	15.00	2.779	2.846	2.4
		2.913		
2	7.50	1.934	1.969	1.8
		2.004		
3	3.75	1.188	1.218	2.4
		1.247		
4	1.88	0.710	0.730	2.8
		0.750		
5	0.94	0.430	0.438	1.9
		0.447		
6	0.47	0.266	0.273	2.6
		0.280		
7	0.23	0.186	0.191	2.8
		0.197		
Blank	0	0.098	0.098	0.4
		0.097		

Table 2

The OD values of the standard curve may vary according to the conditions of assay performance (e.g. operator, pipetting technique, washing technique or temperature effects). Furthermore shelf life of the kit may affect enzymatic activity and thus colour intensity. Values measured are still valid.

## 12. LIMITATIONS

- Since exact conditions may vary from assay to assay, a standard curve must be established for every run.
- Bacterial or fungal contamination of either screen samples or reagents or cross-contamination between reagents may cause erroneous results.
- Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.
- Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Empty wells completely before dispensing fresh wash solution, fill with Wash Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.
- Excessive freeze–thaw cycles may result in loss of bio-active human VEGF-C. Avoid repeated freeze–thaw cycles, preferentially use freshly collected samples.

## 13. PERFORMANCE CHARACTERISTICS

### 13.1 Sensitivity

The limit of detection of human VEGF-C defined as the analyse concentration resulting in an absorbance significantly higher than that of the dilution medium (mean plus 2 standard deviations) was determined to be 0.057 ng/ml (mean of 6 independent assays).

### 13.2 Reproducibility

#### 13.2.1 Intra-assay

Reproducibility within the assay was evaluated in 3 independent experiments. Each assay was carried out with 6 replicates of 6 serum samples containing different concentrations of human VEGF-C. 2 standard curves were run on each plate. Data below show the mean human VEGF-C concentration and the coefficient of variation for each sample (see Table 3). The calculated overall intra-assay coefficient of variation was 6.7%.

Sample	Experiment	Mean Human MMP-9 Concentration (ng/ml)	Coefficient of Variation (%)
1	1	10.46	1.3
	2	10.31	3.3
	3	10.39	5.9
2	1	2.44	2.9
	2	2.16	4.7
	3	2.14	6.8
3	1	1.24	10.3
	2	10.38	11.2
	3	0.92	10.7
4	1	1.14	9.9
	2	0.97	5.1
	3	0.85	10.7
5	1	1.19	3.2
	2	0.89	8.6
	3	0.90	6.0
6	1	0.65	3.2
	2	0.47	10.4
	34	0.47	7.9

Table 3 The mean human VEGF-C concentration and the coefficient of variation for each sample.

### 13.2.2 Inter-assay

Assay to assay reproducibility within one laboratory was evaluated in 3 independent experiments. Each assay was carried out with 6 replicates of 6 serum samples containing different concentrations of human VEGF-C. 2 standard curves were run on each plate. Data below show the mean human VEGF-C concentration and the coefficient of variation calculated on 12 determinations of each sample (see Table 4). The calculated overall inter-assay coefficient of variation was 12.0%.

Sample	Mean Human VEGF-C Concentration (ng/ml)	Coefficient of Variation (%)
1	10.39	0.7
2	2.24	7.5
3	1.07	15.6
4	0.99	14.3
5	0.98	15.1
6	0.53	18.9

Table 4 The mean human VEGF-C concentration and the coefficient of variation of each sample.

### 13.3 Spiking Recovery

The spiking recovery was evaluated by spiking 4 levels of human VEGF-C into serum, plasma and cell culture supernatant. Recoveries were determined in 3 independent experiments with 4 replicates each. The amount of endogenous human VEGF-C in unspiked serum was subtracted from the spike values.

For recovery data see Table 5.

Sample matrix	Spike high (%)	Spike medium (%)	Spike low (%)
Serum	105	100	95
Plasma (citrate)	91	87	87
Cell culture supernatant	96	97	91

Table 5

### 13.4 Dilution Linearity

Serum, plasma and cell culture supernatant samples with different levels of human VEGF-C were analysed at serial 2 fold dilutions with 4 replicates each.

For recovery data see Table 6.

Sample matrix	Recovery of Exp. Val.	
	Range (%)	Mean (%)
Serum	78 - 102	87
Plasma (citrate)	79 - 125	97
Cell culture supernatant	77 - 100	89

Table 6



## 13.5 Sample Stability

### 13.5.1 Freeze-Thaw Stability

Aliquots of serum samples were stored at -20°C and thawed 5 times, and the human VEGF-C levels determined. There was no significant loss of human VEGF-C immunoreactivity detected by freezing and thawing 5 times. Excessive freeze–thaw cycles may result in loss of bio-active human VEGF-C. Avoid repeated freeze–thaw cycles, preferentially use freshly collected samples.

### 13.5.2 Storage Stability

Aliquots of serum samples were stored at -20°C, 2-8 °C, room temperature (RT) and at 37°C, and the human VEGF-C level determined after 24 h. There was no significant loss of human VEGF-C immunoreactivity detected during storage under above conditions.

## 13.6 Specificity

The assay detects both natural and recombinant human VEGF-C. The cross reactivity and interference of circulating factors of the immune system was evaluated by spiking these proteins at physiologically relevant concentrations into a human VEGF-C positive serum. There was no cross reactivity or interference detected, notably not with human VEGF-A, VEGF-B, VEGF-D, PDGF-AA, PDGF-BB, PLGF, HGF, EGF, VEGF-R1 and KDR.

## 13.7 Expected Values

Panels of 40 serum as well as plasma (citrate) samples from randomly selected apparently healthy donors (males and females) were tested for human VEGF-C.

For detected human VEGF-C levels see Table 7.

Sample Matrix	Number of Samples Evaluated	Range (ng/ml)	% Detectable	Mean of Detectable (ng/ml)
Serum	40	nd *- 5.35	90	0.61
Plasma (Citrate)	40	nd *- 0.50	10	0.13

\* n.d. = non-detectable, samples measured below the lowest standard point are considered to be non-detectable.

Table 7

## 14. REFERENCES

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8. Orpana A, Salven P. Angiogenic and lymphangiogenic molecules in hematological malignancies. *Leuk Lymphoma.* 2002 Feb; 43 (2): 219-24.
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Example Version

## 15. REAGENT PREPARATION SUMMARY

### 15.1 Wash Buffer (1x)

Add **Wash Buffer Concentrate 20x** (50 ml) to 950 ml distilled water.

Number of Strips	Wash Buffer Concentrate (ml)	Distilled Water (ml)
1 - 6	25	475
1 - 12	50	950

### 15.2 Biotin-Conjugate

Make a 1:100 dilution of **Biotin-Conjugate** in Conjugate Diluent:

Number of Strips	Biotin-Conjugate (ml)	Conjugate Diluent (ml)
1 - 6	0.06	5.94
1 - 12	0.12	11.88

### 15.3 Streptavidin-HRP

Make a 1:100 dilution of **Streptavidin-HRP** in Conjugate Diluent:

Number of Strips	Streptavidin-HRP (ml)	Conjugate Diluent (ml)
1 - 6	0.06	5.94
1 - 12	0.12	11.88

### 15.4 Human VEGF-C Standard





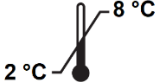




Reconstitute lyophilized **human VEGF-C standard** with Sample Diluent (for subsequent measurement of **serum** or **plasma samples**) or Conjugate Diluent (for subsequent measurement of **cell culture supernatant samples**). (Reconstitution volume is stated in the Quality Control Sheet.)

## 16. TEST PROTOCOL SUMMARY

1. Determine the number of microwell strips required.
2. Wash microwell strips twice with Wash Buffer.
3. Add 80  $\mu$ l Assay Diluent in duplicate to all wells.
4. Add 20  $\mu$ l of prepared Standard dilutions in duplicate to standard wells.
5. Add 20  $\mu$ l sample in duplicate, to designated sample wells.
6. Add 20  $\mu$ l of Sample Diluent (for serum or plasma samples) or Conjugate Diluent (for cell culture supernatant samples) in duplicate to blank wells.
7. Cover microwell strips and incubate 2 hours at room temperature (18° to 25°C).
8. Prepare Biotin-Conjugate.
9. Empty and wash microwell strips 6 times with Wash Buffer
10. Add 100  $\mu$ l Biotin-Conjugate to all wells.
11. Cover microwell strips and incubate 1 hour at room temperature (18° to 25°C).
12. Prepare Streptavidin-HRP.
13. Empty and wash microwell strips 6 times with Wash Buffer.
14. Add 100  $\mu$ l diluted Streptavidin-HRP to all wells.
15. Cover microwell strips and incubate 1 hour at room temperature (18° to 25°C).
16. Empty and wash microwell strips 6 times with Wash Buffer.
17. Add 100  $\mu$ l of TMB Substrate Solution to all wells.
18. Incubate the microwell strips for about 30 minutes at room temperature (18° to 25°C).
19. Add 100  $\mu$ l Stop Solution to all wells.
20. Blank microwell reader and measure colour intensity at 450 nm.

**Note:** If instructions in this protocol have been followed, samples have not been diluted and the concentration read from the standard curve must not be multiplied by a dilution factor.

## 17. EXPLANATION OF SYMBOLS

	Catalogue number
	Batch code
	Caution
	Use by date
	Temperature limit
	Manufacturer
 <p data-bbox="260 1184 467 1216">www.biovendor.com</p>	Read electronic instructions for use - eIFU
	The content is sufficient for 96 tests
	Biological risks





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